

Instruction for use

Serotonin ELISA

Enzyme Immunoassay for the Quantitative Determination of Serotonin in Serum, Plasma and Urine

IVD CE

REF EA602/96

 $\sqrt{\Sigma}$ 12 x 8

±2√ ⁺⁸ 2 − 8 °C

DLD Gesellschaft für Diagnostika und medizinische Geräte mbH Adlerhorst 15 • 22459 Hamburg • Tel 040-555 87 10 • Fax 040-555 87 111 Internet: http://www.dld-diagnostika.de • E-Mail: contact@dld-diagnostika.de

ser-e_4.doc 2020-02-21

Contents

1.	Introduction and Principle of the Test	Page	3
2.	Precautions	Page	3
3.	Storage and Stability	Page	3
4.	Contents of the Kit	Page	4
5.	Sample Collection	Page	5
6.	Preparation of Reagents and Samples	Page	6
7.	Test Procedure	Page	7
8.	Calculation of the Results	Page	8
9.	Assay Characteristics	Page	9
10.	Literature	Page	11
	Pipetting Scheme	Page	12

Symbols

CE EC Declaration of Conformity IVDIn Vitro Diagnostic Medical Device CONT Content **Expiry Date** Temperature limitation LOT Batch code Sufficient for ... determinations Manufacturer $\bigcap_{\mathbf{i}}$

Consult instructions for use

Hazard Pictograms



REF

Danger

Catalogue number



1. Introduction and Principle of the Test

Serotonin (5-Hydroxytryptamine), a biogenic amine, is a product of the tryptophan metabolism. It is a well evaluated neurotransmitter of the central nervous system and can be found in high concentrations in the chromaffine cells of the intestinal mucosa, in the platelets and the serotonergic neurones of the brain.

Central-serotonergic neurones influence physiological functions such as sleep and the hormonal and cardio-vascular regulation. Increased serum levels can be found with malignant carcinoid, endogenous depression and schizophrenia.

The assay kit provides materials for the quantitative measurement of derivated serotonin (5-Hydroxytryptamine) in serum, plasma and urine. The derivation is performed during the preparation of the samples. By using the acylation reagent the serotonin is quantitatively derivated into N-acylserotonin.

The competitive Serotonin ELISA kit uses the microtitre plate format. Serotonin is bound to the solid phase of the microtiter plate. Acylated serotonin and solid phase bound serotonin compete for a fixed number of antiserum binding sites. When the system is in equilibrium, free antigen and free antigen-antiserum complexes are removed by washing. The antibody bound to the solid phase serotonin is detected by antirabbit/peroxidase. The substrate TMB / peroxidase reaction is monitored at 450 nm. The amount of antibody bound to the solid phase serotonin is inversely proportional to the serotonin concentration of the sample.

2. Precautions

- For in vitro diagnostic use only.
- Disposable gloves and safety glasses should be used.
- Material of animal origin used in the preparation of the kit has been obtained from animals certified as healthy but these materials should be handled as potentially infectious.
- Some components of this kit are containing hazardous reagents.
 These components are marked with the adequate hazard label.

3. Storage and Stability

On arrival, store the kit at 2-8 °C. Once opened the kit is stable until its expiry date. For stability of prepared reagents refer to Preparation of Reagents.

Do not use components beyond the expiration date shown on the kit labels.

Do not mix various lots of any kit component within an individual assay.

4. Contents of the Kit

for acylation

STRIPS 4.1 **MT-Strips** 12 strips 8 wells each, break apart, precoated with serotonin CAL 1-6 4.2 Standards 1 - 6 6 vials Each 4 ml, ready for use Concentrations: Standard 1 2 3 4 5 6 150 500 ng/ml 0 15 50 2,500 4.3 Control 1 & 2 CONTROL 1 & 2 2 vials Each 4 ml, ready for use, Range: see g.c. certificate ACYL-BUFF 4.4 **Acylation Buffer** 1 vial 3 ml, ready for use, blue coloured Warning ACYL-REAG 1 vial 4.5 **Acylation Reagent** 2.5 ml, ready for use Danger, Warning 4.6 Antiserum AS 1 vial 11 ml, ready for use, colour coded yellow Rabbit-anti-N-acylserotonin 4.7 CONJ **Enzyme Conjugate** 1 vial 12 ml, ready for use, Goat anti-rabbit-IgG-peroxidase WASH Wash Buffer 4.8 1 vial 20 ml, 50x concentrated Dilute contents with dist, water to 1 litre total volume. 4.9 **Substrate** SUB 1 vial 12 ml TMB solution, ready for use 4.10 Stop Solution STOP 1 vial 12 ml, ready for use Contains 0.3 M sulphuric acid **ACYL-PLATE** 4.11 Reaction plate 1 piece

1 vial

lyophilzed, dissolve content with 20.5 ml dist. water, dissolve carefully to minimize foam formation

Additional materials and equipment required but not provided:

- Pipettes (10, 25, 50, 100 and 200 µl)
- Orbital shaker
- Microplate washing device
- Microplate photometer (450 nm)

5. Sample Collection

5.1. Serum and Plasma

The test can be performed with serum as well as with EDTA plasma. If plasma is to be used care must be taken to get true platelet-free plasma. Otherwise, the Serotonin level has to be related to the number of thrombocytes in the sample. Since the preparation of platelet-free plasma requires special precautions, it is generally recommended to use serum instead of plasma.

Hemolytic and lipemic samples should not be used.

The samples can be stored up to 6 hours at 2 - 8 °C. For a longer storage (up to 6 months) the samples must be frozen at -20 °C

Repeated freezing and thawing should be avoided.

5.2. Urine

The total volume of urine excreted during a 24-hours period should be collected and mixed in a single bottle containing 10 - 15 ml of 6 M hydrochloric acid as preservative. Avoid exposure to direct sun light. Determine the total volume and take an aliquot for the measurement. For patients with suspected kidney disorders the creatinine concentration should be tested, too. Urine samples can be stored at -20 °C for at least 6 months.

6. Preparation of Reagents and Samples

6.1. Microtiter strips STRIPS

Before opening the packet of strip wells, allow it to stand at room temperature for at least 10 minutes. After opening, keep any unused wells in the original foil packet with the desiccant provided. Reseal carefully and store at 2-8 °C.

6.2 Wash Buffer WASH

Dilute the content with dist. water to a total volume of 1,000 ml. For further use the diluted wash buffer must be stored at 2 - 8 °C for a maximum period of 4 weeks.

6.3. Equalizing Reagent **EQUA-REAG**

Dissolve the content with 20.5 ml dist. water, mix shortly and leave on a roll mixer for 30 minutes. Handle carefully in order to minimize foam formation. The reconstituted Equalizing Reagent should be stored frozen at -20 °C and is stable for a minimum of 1 year.

All other reagents are ready for use.

6.4. Preparation of Samples (Acylation)

The wells of the reaction plate for the acylation can be used only once. Please mark the respective wells before using.

- 1. Pipette each 20 μl standard 1 6, each 20 μl control 1 & 2, each 20 μl serum, 20 μl urine or 40 μl plasma into the respective wells of the reaction plate.
- 2. Pipette each 20 μl Acylation Buffer into all wells.
- 3. Pipette each 200 µl Equalizing Reagent into all wells Shake plate on an orbital shaker for 10 seconds.
- 4. Pipette each 20 μl Acylation Reagent into all wells, mix immediately. Please note that Acylation Reagent reacts with many plastic materials including plastic trays. It does not react with normal pipette tips and with glass devices. Use an Eppendorf multipette or similar, fill the syringe directly from the vial and add well by well.
- 5. Incubate for 15 minutes at room temperature (approx. 20 °C) on an orbital shaker. Colour changed to green.
- 6. Take each 20 µl for the ELISA.

7. Test Procedure

7.1 Sample Incubation

Pipette each 20 µl prepared Standards 1 to 6, 20 µl prepared controls and 20 µl prepared samples into the respective wells of the coated microtiter strips (duplicates are recommended).

Pipette each 100 µl Antiserum into all wells.

Incubate for 30 minutes at room temperature on an orbital shaker.

7.2 Washing

Discard or aspirate the contents of the wells and wash thoroughly with each 250 µl Wash Buffer. Repeat the washing procedure 3 to 4 times. Remove residual liquid by tapping the inverted plate on clean absorbent paper.

7.3 Conjugate Incubation

Pipette each 100 µl enzyme conjugate into all wells. Incubate for 15 minutes at room temperature on an orbital shaker.

7.4 Washing

Repeat step 7.2.

7.5 Substrate Incubation

Pipette each 100 μ l Substrate into all wells and incubate for 15 \pm 5 minutes at room temperature on an orbital shaker.

7.6 Stopping

Pipette each 100 µl Stop Solution into all wells.

7.7 Reading

Read the optical density at 450 nm (reference wavelength between 570 and 650 nm) in a microplate photometer.

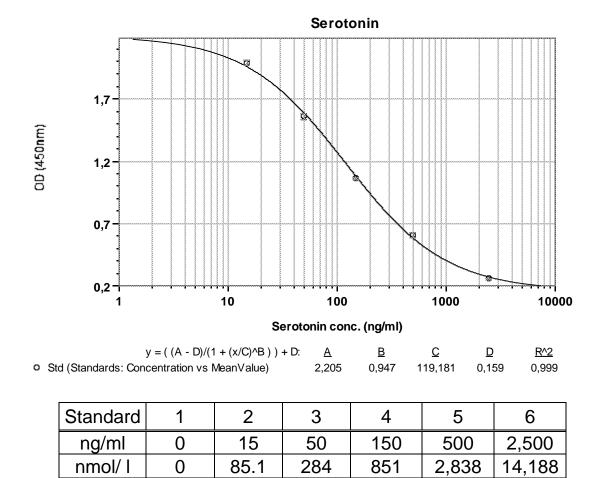
8. Calculation of the Results

On a semilogarithmic graph paper the concentration of the standards (x-axis, logarithmic) are plotted against their corresponding optical density (y-axis, linear). Alternatively, the optical density of each standard and sample can be related to the optical density of the zero standard, expressed as the ratio OD/OD_{max}, and then plotted on the y-axis.

The concentration of the controls, urine and serum samples can be read directly from this standard curve by using their average optical density.

The read-off values for plasma samples have to be divided by a factor of 1.8.

Typical standard curve:



Conversion: Serotonin: 1ng/ ml = 5.675 nmol/l

9. Assay Characteristics

Normal Range

The reference ranges given below should only be taken as a guideline. It is recommended that each laboratory should establish its own normal values.

Plasma (platelet-free): < 10 ng/ml

Serum: Female 80 - 450 ng/ml

Male 40 - 400 ng/ml

Urine: 50 - 250 μg/day

Sensitivity

4.7 ng/ml for serum and urine

2.6 ng/ml for plasma

Specificity (Cross Reactivity)

Structural related components were tested for possible interference with the antisera against Serotonin used in the ELISA method. The tested compounds were Tryptamine, Melatonin, 5-HIAA, 5-Hydroxy-L-Tryptophan, 5-Methoxytryptamine and L-Tryptophan.

Substance	ED-50-Value (ng/ml)	Cross Reactivity (%)
Serotonin	133	100
Tryptamine	8,700	1.5
5-Methoxytryptamine	56,900	0.23
Melatonin	> 1,000,000	< 0.0133
5-Hydroxy-L-Tryptophan	> 1,000.000	< 0.0133
5-HIAA	> 10,000,000	< 0.00133
L-Tryptophan	> 10,000,000	< 0.00133

Recovery

Increasing amounts of Serotonin were added to a serum, plasma and an urine sample. Each spiked sample was assayed. The analytical recovery of Serotonin was estimated at different concentrations by using the theoretically expected and the actually measured values.

Matrix	Range (ng/ml)	Mean (%)	Range (%)
Serum	70 - 824	95	85 - 105
Urine	27 - 1085	105	83 - 120
Plasma	62 - 293	96	87 - 102

Linearity

The linearity of the ELISA method was investigated using different dilutions of a serum, plasma and an urine sample. Samples were diluted with distilled water.

Matrix	Range (ng/ml)	max. Dilution	Mean (%)	Range (%)
Serum	60 – 1,203	1 : 20	91	83 - 97
Urine	66 – 1,316	1 : 20	100	96 - 104
Plasma	79 - 395	1:5	94	90 - 97

Reproducibility

The reproducibility of the ELISA method was investigated by measuring the intra-assay-coefficients of variation (cv) by repeated measurements of different serum, plasma and urine samples with different Serotonin concentrations.

Intra-Assay Variation

Matrix	Range (ng/ml)	cv (%)
Serum	148 – 497	7.3 - 6.9
Urine	93 – 209	6.7 – 6.1
Plasma	163	7.7

10. Literature

- Kema, P.; de Vries, E.; Muskiet, F. (2000):
 Clinical chemistry of serotonin and metabolites
 Journal of Chromatography B, 747 33–48
- Lechin, F.; van der Dijs, B.; Lechin, A. (2005):
 Circulating Serotonin, Catecholamines, and CentralNervous System Circuitry
 Relatedto Some Cardiorespiratory, Vascular, and HematologicalDisorders
 The Journal of Applied Research Vol. 5, No. 4
- Spaeth, M. (2006):
 Fibromyalgia Syndrome: The Role of Neurochemicals
 Primary Psychiatry. 13(9):72-75
- Spivak, B.; Vered, Y.; Graff, E.;. et al. (1999):
 Low Platelet-Poor Plasma Concentrations of Serotoninin Patients with Combat-Related Posttraumatic Stress Disorder
 BIOL PSYCHIATRY, 45:840–845
- Ernberg, M.; Lundeberg, T.; Kopp, S. (2000):
 Pain and allodynia/hyperalgesia induced by intramuscular injection of serotonin in patients with fibromyalgia and healthy individuals
 Pain 85 31±39
- Alvarez, J.; Gluck, N.; Fallet, A.; et al. (1999):
 Plasma serotonin level after 1 day of fluoxetine treatment: a biological predictor for antidepressant response?
 Psychopharmacology 143: 97.101
- Mück-Seler, D.; Pivac, N.; Jakovljevic, M.; et al. (1999):
 Platelet Serotonin, Plasma Cortisol, and Dexamethasone Suppression Test in Schizophrenic Patients
 BIOL PSYCHIATRY 45:1433–1439
- Vikenes, K.; Farstad, M.; Nordrehaug, J. (1999):
 Serotonin Is Associated with Coronary Artery Disease and Cardiac Events Circulation August 3, 1999; 483-489
- Dayan, P.; Huys, Q. (2008):
 Serotonin, Inhibition, and Negative Mood
 PLoS Computational Biology February 2008 | Volume 4 | Issue 2 | e4
- Leboyer, M.; Philippe, A.; Bouvard, M.; et al. (1999):
 Whole Blood Serotonin and Plasma Beta-Endorphin in Autistic Probands and Their First-Degree Relatives
 BIOL PSYCHIATRY 1999;45:158–163

Pipetting Scheme

Sample Preparation

		Standards	Control	Serum, Urine	Plasma
Standard 1 - 6	μl	20			
Control 1 & 2	μl		20		
Serum, Urine	μl			20	
Plasma	μl				40
Acyl. Buffer	μl	20	20	20	20
Equalizing Reag.	μl	200	200	200	200

Shaking the plate for 10 seconds on an orbital shaker

Acyl. Reagent	ul	20	20	20	20

mix Immediately and incubate 15 minutes at room temperature on an orbital shaker

Take each 20 µl for the ELISA.

Pipetting Scheme ELISA

		Standard	Control	Sample
Standard 1 - 6	μl	20		
Control 1 & 2	μl		20	
Sample	μl			20
Antiserum	μl	100	100	100

30 minutes incubation at room temperature on an orbital shaker

3 - 4 x washing

Conjugate	μl	100	100	100

15 minutes incubation at room temperature on an orbital shaker

3 - 4 x washing

Substrate µl 100 100 100

15 ± 5 minutes incubation at room temperature on an orbital shaker

Stop Solution	μl	100	100	100

Reading of absorbance at 450 nm